

University of Vermont. Unfortunately, none of the First Detector award recipients were present to receive their awards but they were notified and sent their plaques.

The NPDN Outstanding Service Award was presented to an individual that performed outstanding service and gone above and beyond the call of duty. The first individual award was presented to Rachel McCarthy as editor of the *NPDN News*. The second individual award was presented to Sara May of the Pennsylvania State University for her leadership in coordinating the NPDN-PSU Basic Techniques and Fusarium Identification Workshops. The third individual award was presented to Gail Ruhl of Purdue University for her many contributions to the NPDN. The fourth individual award was presented to Carla Thomas of the University of California at Davis for her many contributions to the NPDN.

We concluded the evening with a fun award...the newly created NPDN Rotten Tuber Award. This award was formed to give a stage for NPDN members and/or colleagues to showcase those very

unique and/or unusual samples that create stories worth sharing within our community. The award submissions were evaluated based on the uniqueness of the situation, the story telling ability of the submitter, and the shock value. The 3<sup>rd</sup> place award was presented to Connie Tande (Picture I) of South Dakota State University for her submission about a suspect fungal growth (Picture K) found on the side of a road. The 2<sup>nd</sup> place award was presented to Cheryl Smith (Picture J) of the University of New Hampshire for her submission of "linoleum blight". The 1<sup>st</sup> place award was presented to Ruth O'Neill (no picture available) of Montana State University for her submission of an ant named Gigi.

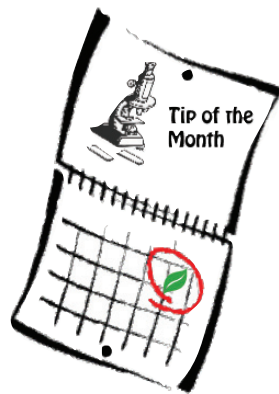
The NPDN awards presentation and banquet at the third National Meeting were a lot of fun and we appreciate everyone's contributions to make it a huge success! If you would like more details about the awards and those who received them this year, please visit the 2011 National Meeting website at [www.npdn.org/node/119](http://www.npdn.org/node/119). And remember to save those unique samples for the fourth National Meeting! 🌿

## Diagnostic Updates

### Use of Autoclaved Pine Needles for Inducing Sporulation of *Botryosphaeria* Isolated from Plant Tissue

Compiled by Gail Ruhl; Purdue Plant and Pest Diagnostic Lab, Purdue University

I recently attended an excellent hands-on *Botryosphaeria* identification workshop



presented by Dr. Jose Ramon Urbez-Torres, Dr. Suzanne Rooney-Latham and Dr. Cheryl Blomquist at the NPDN Third National Meeting. We were given the opportunity to examine many different cultures and practice correct identification of species using conidial morphology (size, shape, pigmentation, septation, etc.). We learned that formation of pycnidia and conidia of some of the *Botryosphaeria* species, such as *Fusicoccum* and *Neofusicoccum* do not always occur on regular culture media and under standard growing conditions. Special techniques such as the use of a host substrate (e.g. pine needles) on a low nutrient media, (e.g. water agar) may be necessary to induce

production of pycnidia and conidia. Twice autoclaved pine needles are placed on the surface of water agar and small mycelial plugs from suspect cultures are

## NPDN/USDA-APHIS 2012 Advanced Diagnostic Workshops

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The NPDN Diagnostics Program Area Committee and members of USDA-APHIS-PPQ-CHPST-National Plant Germplasm and Biotechnology Laboratory (NPGBL) are pleased to announce the 2012 advanced diagnostic workshops. During the spring of 2012, we will offer advanced trainings on a number of topics. One of the most important sessions offered is on citrus leprosis (CiLV) to include sweet orange scab (SOS) and citrus black spot (CBS). We are hoping diagnosticians from all the states where citrus is grown will attend this 4½ day session. The training will cover disease symptoms and methods of detection and identification of CiLV, CBS and SOS. The molecular diagnostics will include PCR, real-time PCR, RT-PCR and RT-real-time PCR. Another topic offered will be bioinformatics, part one (1½ day session), which has been offered previously and will cover analysis of obtained sequences from both plus and minus strands, editing sequences, blasting sequences, understanding blast results based

on size and gene target, when to directly sequence PCR products or clones, which genes are used for sequence analysis for fungi, bacteria, and viruses, what sequence analysis programs are available commercially or as freeware, and hands-on use of sequence analysis programs using sequences from case studies for different pathogen types, and bioinformatics part two (1½ day session), which is a new addition to the line-up and will allow the participants to work with their own sequences. Due to continued interest, we have scheduled three *Phytophthora* training sessions. The

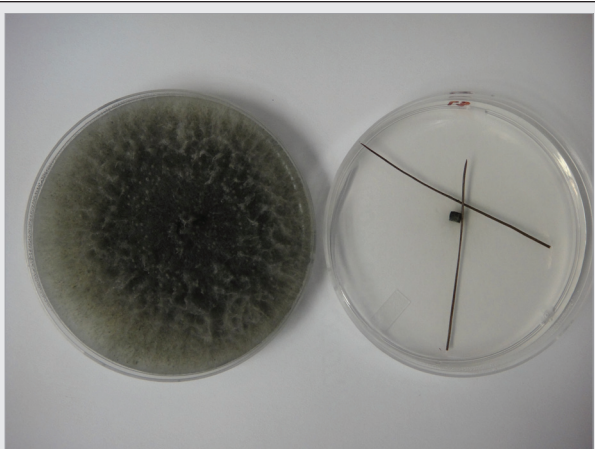


Figure 1. Plug from nonsporulating culture placed on 'pine needle' agar. Photo courtesy of S. Latham.



Figure 2. Pine needles. Photo courtesy of Nancy Harding University of Maryland.

then placed on the media (mycelium side down) next to the pine needle (Figures 1 and 2). Pycnidia usually develop within a month.

### Useful websites for fungal ID:

[www.crem.fct.unl.pt/botryosphaeria\\_site/](http://www.crem.fct.unl.pt/botryosphaeria_site/)

<http://nt.ars-grin.gov/fungal databases/fungushost/fungushost.cfm>

[www.mycobank.org/](http://www.mycobank.org/) 